

Assessment of Some Maize Varieties for Their Tolerance and Suppression to *Sitophilus zeamais* (mostoch), (Coleoptera: *Curculionidae*)

AMINU M. A¹, NAMADINA M.M², HAFIZU M. S³, AHMAD A. T⁴

¹Department of Biological Sciences, Bayero University Kano-Nigeria

²Department of Plant Biology, Bayero University Kano-Nigeria

^{3,4} Department of Biological Sciences, Federal University of Kashere, Gombe State Nigeria.

Abstract - Insect pests cause major damage to stored maize grain thereby reducing its weight, quality and germination vigour. Five open pollinated maize varieties (YLW-123, RED-COB, Single Cross, Hybrid-White, Local-I, and Local-II) and one hybrid maize variety (YLW-123) were evaluated for tolerance and their effects on progeny development against the maize weevil, *Stophilius zeamais*. The experiment was laid in a randomized complete block design, with 6 treatments replicated 5 times. 100g maize grain was infested with 100 three-week-old unsexed pure culture adult weevils in 750 ml jars. After 15 days oviposition period, adult weevils were sieved out and parent weevil mortality determined. After a further 45 days, number of weevils emerged, percentage gain weight loss and number of damaged kernels were determined. Percentage kernel germination was determined through a germination test after 45 days of weevil attack. There were significant differences ($p < 0.05$) in number of parent weevil mortality, number of weevils emerged, grain weight loss, kernel damaged and germination percentage among varieties. YLW-123 and RED-COB varieties showed potential to *S. zeamais* progeny suppression and tolerance as evidenced by high parent weevil mortality, low weevil emergence, less grain weight loss, low grain damage and high germination percentage.

Indexed Terms -*Stophilius zeamais*, Maize Varieties, Tolerance, Suppression.

I. INTRODUCTION

Maize (*Zea mays* L.) is the most important crop in Nigeria since it is a staple food crop which is widely grown by most smallholder farmers who significantly contribute to national production [1]. The necessity to increase maize production cannot be over emphasized; in Nigeria it ranks second after rice in terms of total cereal production, number of producers and area grown

[2]. It has been reported that the crop accounts for 70% of the total hectare under cereals with 60% of the whole production coming from the small-scale farmers [3]. *Sitophilus zeamais* is a serious pest of economic importance in stored products worldwide [4]. The pest is so devastating and is capable of multiplying to large populations causing tremendous damage to the grain [5]. It is estimated that about 10– 40% of the total damage to stored grains worldwide is caused by insect pests [6] of which they account for approximately 5– 10% of maize grain loss in Southern Africa [7]. Under severe infestations, maize weevils can cause up to 90% loss of stored grain [8]. Grain weight loss of 12–20% and 80% caused by the maize weevil is common in untreated maize grain stored in traditional structures in tropical countries [9]. It has been reported that much of the maize produced by the smallholder farmers in Nigeria is lost to weevil attack and very little research has been done on the development of affordable alternatives which offer same control levels to weevils as pesticides [10]. Although synthetic pesticides can control it, majority of communal farmers are resource-poor and have no means and proper skills to acquire and handle them. Moreover, pesticides are expensive, not readily available and pose health problems to consumers due to their toxicity since many have some residual effect. Evidence from different African countries illustrates that improper use of chemicals is causing loss of life and negative repercussions on human health [11] and other problems associated with their use are loss of efficacy, regulatory restrictions because of adverse effect on non-targeted organisms and eco-toxicity [12].

Decrease in agricultural productivity exposes local farmers and the nation to chronic food shortages hence

it is a serious threat to mankind. This creates the need for farmers to come up with mechanisms for conserving their scarce food resource base. The constraints to maize seed availability and affordability have prompted the need to sow open pollinated varieties (OPVs) by resource constrained smallholder farmers in Nigeria. The advantages to the use of insect resistant varieties are especially important in developing countries where farmers can rarely afford to purchase insecticides for crop protection [13]. These varieties provide practical and economic way to minimize field and grain storage losses to improve both quantity and quality of stored grain for planting and human consumption (16) Insect resistant crops greatly increase farming efficiency by reducing or eliminating the costs of insecticides and the risk of yield losses from insect damages. Grain resistance as a method of pest control is advantageous since most resistant varieties maintain high levels of resistance for a long time despite upsurge of biotypes [14]. The potential negative effects associated with insecticide use are eliminated with the use of insect resistant varieties. In many developing countries, the demand for maize surpasses that for other food crops due to the growth in meat and poultry consumption, which consequently, have led to the rapid increase in the demand for maize as livestock feed. Thus, there is a need to develop cheaper, equally effective and safer alternatives for insect pest control, including host plant tolerance [15]. However, the level of OPVs resistance or tolerance to weevil attack is not fully understood hence there is need for screening of maize grain varieties for maize weevil evaluations. Considering the economic importance of maize in the country as well as the destructive nature of *S. zeamais* to the crop, the present study was undertaken with the main objective of screening different stored maize varieties grain for tolerance against the maize weevil.

II. MATERIALS AND METHODS

The experiment was conducted at biological Science laboratory, Department of Biological science, Faculty of life science Bayero University Kano. The study location is Bayero University, Kano city, in Kano state Nigeria. It is situated between latitudes 9.6° 25' N to 6.8°47' N and longitude 8° 22' E to 8°39' E and 472m above sea level. Bayero University, Kano (Old Campus) is found in Gwale LGA bordered by

Ungoggo, Dala and Fagge Local Government Areas (LGA) to the South West.

Six pure maize pollinated varieties namely, YLW-123, RED-COB, Single Cross, Hybrid-White, Local-I, and Local-II were used in the experiment. A randomized complete block design was used to arrange the jars in the laboratory to minimize the door effects and each treatment was replicated five times. The maize grain was thoroughly cleaned using a 1 mm sieve-mesh screen so as to remain with grain with intact testate, which was then disinfested by keeping it in a deep freezer at -4°C for 2 weeks. The moisture content of the grain was in the range of 12-13%. Approximately 100 g of each of the maize varieties was placed into the 750 ml jars with perforated lids. One hundred three -week old unsexed adult weevils were introduced into each jar. The jars were placed in the shelves at a temperature range of $28\pm 2^{\circ}\text{C}$ and relative humidity of $70\pm 5\%$. Weevils used in the experiment were obtained from the Crop Protection Department at Audu Bako College of Agriculture, Dambatta. Grain from the previous season was used to prepare the pest culture. Grain was first sieved to remove dirt and broken particles. Three 750 ml consul jars with perforated lids to allow for air circulation were filled with grain to the three-quarter level. Filter paper was put inside each of the perforated lids to prevent insects from escaping. The jars containing the grain were placed in a freezer for 2 weeks to kill any insect eggs which might have been present in the grain. The grain was then transferred into the shelves and stored for 3 weeks to achieve uniform grain temperature and moisture content. The temperature was set at $31\pm 4^{\circ}\text{C}$ and humidity at $75\pm 5\%$. After 3 weeks each consul jar was infested with 100 adult weevils and the jars were placed in the shelves. After 14 days oviposition period the grain was sieved to discard adult weevils which had laid eggs in the kernels. Maize weevils take about 30 days to complete their life cycle [16], so after 30 days the weevils began to emerge. After 35 days the F1 progeny was collected by sieving damaged grain. The adult weevils collected were in the range of 1–3 days old. The weevils were later used in the evaluation of OPV's for weevil resistance. A refrigerator was used for disinfestations of seed by storing the seed at -4°C for two weeks. Camel hair brush was used for collecting insects and the tweezers for holding the insects. A 1 and 4.7 mm screen meshes were used for

separating grain, dust and insects. Weevil mortality was assessed 14 days after the introduction of the insects. The grain was sieved and the number of dead and live pests was counted from each jar to obtain parent weevil mortality. The following formula was used to calculate the percentage weevil mortality.

$$\text{Weevil mortality} = \frac{\text{number of dead pests}}{\text{total number of pests}} \times 10$$

After 14 days, the weevils which emerged from the grain in each jar were counted and their number was recorded. After 45 days of incubation the grain was sieved, dust removed and the clean grain was weighed and expressed as a percentage weight loss of the original weight [17]. Five days after incubation, the grain was thoroughly mixed and 30 maize kernels (grains) were randomly selected to assess the level of grain damage. The grain was sorted into damaged (grain with holes and/or tunnels) and undamaged grain. Grain in each fraction was counted and the number of damaged grain recorded. Maize grain genotypes exposed to maize weevils for 45 days was germinated in an incubator at a temperature of 28°C in Petri- dishes in moist wrapping papers. Twenty seeds per maize grain genotype were placed on top of the moist paper in Petri- dishes. The Petri- dishes were covered and put into an incubator for 10 days at 28°C. Germination percentage was calculated using the formula;

$$\text{Germination \%} = \frac{G1}{G2} \times 100; \text{ Where } G1 = \text{total germinated grain, } G2 = \text{total grain in Petri -dish}$$

A. Statistical Analysis

A general analysis of variance (ANOVA) for parent weevil mortality, number of weevils emerged, percentage grain weight loss, kernel damage and percentage germination was conducted using GenStat statistical package version 12. Mean separation was done by using least significant difference (LSD) to compare the significant differences between the treatments at 5% level of significance.

III. RESULT AND DISCUSSION

Weevil attack high parent weevil mortality may also be due to antixenosis, that is, resistance mechanisms which deter colonisation by the insect [18]. High parent weevil mortality might also be attributed to

absence of nutritional factors in the grain which might be important for insect development [19]. Hybrid-White, YLW-123 and Local-I had the lowest parent weevil mortality indicating high susceptibility to weevil attack.

B. Number of Weevil Emergence

There were variations and significant differences ($p < 0.05$) were observed among the varieties in the number of weevils which emerged. The hybrid-White had the highest number of weevils which emerged followed by YLW-123 and Local-II whilst Single Cross and Red-Cob had the least. The mean number of weevils emerged ranged from 0.6-16.2 (Figure 2). The differences in the number of weevils emerged showed that there existed variation in susceptibility to maize weevil attack among the varieties. The varieties which recorded the highest number of weevils emerged indicated greatest susceptibility to maize weevil attack and this might have been due to lack of resistance mechanisms in or on the grain [20]. The low weevil emergence in varieties YLW-123 and Local-II can be attributed to high mortality of parent weevils. These parent weevils might have died before laying eggs or after laying few eggs thus few progeny resulted. The low weevil emergence in these varieties may possibly be attributed to absence of essential nutrients and unbalanced proportion of nutrients leading to the death of the larvae [21]. The significant variation for number of weevils emerged among the varieties could be due to antibiosis effects in resistant varieties leading to retarded development of weevil progeny and sometimes death of weevils before laying eggs [22].

Highest parent weevil mortality followed by YLW-123, Local I and Hybrid- White while Local II had the least parent weevil mortality. Mean percentage parent weevil mortality ranged from 7.6–24.9 (Figure 1). The highest mortality which was observed in Single-Cross and Red-Cob variety could be due to physical factors such as antibiosis or hardness as a result of biochemical compounds which are toxic to the insects which led to subsequent death of the weevils [23]. This indicated that these two varieties have resistant factors in or on their grain which helped to prevent damage.

C. Grain Weight Loss and Damage

Maize grain weight loss and damage were highly significant ($p < 0.001$) among the experimental varieties. Single-Cross recorded the highest weight loss, followed by Local II, Single-Cross, and Local I varieties had the lowest weight loss (Table 1). Local-II had the highest number of damaged grain after 45 days exposure maize weevil followed by Single-Cross, Hybrid-White, YLW-123 had the least number of damaged grains. The mean number of damaged grain ranged from 1.23 –9.12 (Table 1). The researcher considered weight loss and grain damage as the most indicators of a variety's susceptibility to weevil attack. Low weight loss in Local II and YLW123 could be due to resistance mechanisms in or on the grain which prevented weevil attack. Single-Cross had the greatest weight loss thus could be said to be more susceptible to weevil attack than other experimental varieties. Resistance mechanisms could be in the form of deterrents which could be biochemical or morphological or a combination of both [24]. Biochemical compounds in the form of phenolic amides such as defeuroyl and dicoumaroyl may be antibiosis factors to the *S. zeamias* [25]. These phenolic compounds have been detected by fluorescence imaging techniques which clearly show the phenolic barrier to insects in the outer tissue [26]. It has also been reported that antibiotic effects increased restlessness of insects which reduced feeding and could explain how grain damage and weight loss were low among resistant varieties [27]. Some researchers [28] also suggested that variation in maize hybrids was due to antibiosis. Less grain damage could be attributed to antixenosis mechanisms like a smooth pericarp which could deter weevils from oviposition and feeding and also prevents mandibles from gripping maize kernels. The great variation observed in the germplasm evaluated forms a genetic resource base for further improvement to raise the levels of resistance to *S. zeamias* while conserving the farmer preferred traits. This variation in response to the maize weevil attack gives is evident of genetic diversity existence hence a rich genetic resource base for breeding for resistance exists. This offers the opportunity to exploit the variability with the aim of reducing post-harvest insect-pest losses through genetic improvement [29]. This implies that most of

the variation among the genotypes is due to their genetic make-up with little influence from the environment, suggesting that maize improvement for resistance to storage pests is possible through selection.

Table 1. Mean percentage grain weight loss and grain damage among different varieties after 45 days of exposure to *S. zeamias*

variety	% grain weigh loss	No. of Damage grain
YLW-123	0.12	1.24
RED-COB	0.03	0.41
HYBRID-WHITE	2.33	12.61
Single-Cross	7.31	16.0
Local I	5.13	12.00
Local II	4.9	9.12

Table 2 Mean Emerged number of *Stophilius zeamias* on six varieties before and after infection

Maize variety	Initial weevil count	Final weevil count
YLW-123	10	83
RED-COB	10	43
HYBRID-WHITE	10	90
Single-Cross	10	28
Local I	10	22
Local II	10	18

D. Germination

Significant differences ($p < 0.05$) were observed among the treatments. Percentage germination after weevil attack was highest in variety YLW-123 followed by Single-Cross, RED-COB, Local II and Local I while Hybrid-White had the least percentage germination. Mean germination percentage ranged from 68–96% with a mean of 80.3% (Figure 3). The observed differences in germination percentages showed that the varieties differed in susceptibility to maize weevils. Single-Cross and Local II had the highest germination percentage indicating high ability to germinate after exposure to maize weevils. These

varieties also recorded the least number of weevils emerged, highest mortality and least grain weight loss. Thus, these two varieties might have resistance factors which could result in less maize weevil damage thus ability of the grain to germinate is not affected much by maize weevil attack. YLW-123, Hybrid-White and Local I varieties had low germination percentages indicating their susceptibility to maize weevil. This might be due to lack of resistance mechanisms within or in the grain to protect it from weevil attack. Weevil damaged grain germinated and this might be attributed to the fact that the weevils did not damage the embryo.

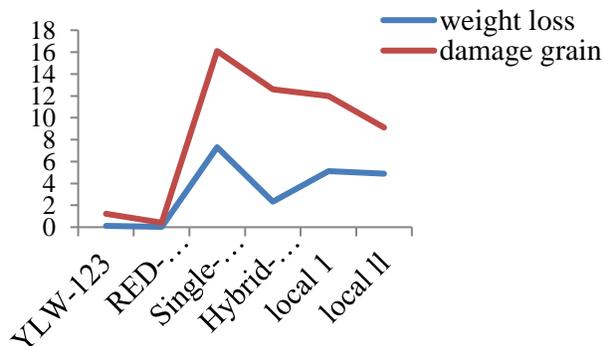


Figure 1. Percentage number of *Sitophilus zeamais* on maize varieties

IV. CONCLUSION

The investigation showed that varieties had different response to maize weevil attack from very susceptible, moderately to tolerance. YLW -123 and RED-COB were highly tolerant as evidenced by the least weight loss, grain damage, number of weevil emerged and highest parent weevil mortality and kernel germination. Single-Cross and Local II had moderate tolerant while Local II and Hybrid-White were highly susceptible. Breeding programme should aim at breeding YLW-123 and RED-COB maize weevil tolerant grain since there is evidence that some tolerant factors exist in the gene pool and ensure food security at family household level.

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REFERENCES

- [1] Rukuni M, Tawonezvi P and Eitche C. Zimbabwe's Agricultural Revolution Revisited, University of Zimbabwe Publications, Harare, 2006, pp 119-140.
- [2] Rukuni M and Eitche CK. 1994. Agricultural Revolution. University of Zimbabwe Publications. Pp 208-212.
- [3] Masoka JK. 2002. Maize production facts and trends in Zimbabwe. University of Zimbabwe Publishers, Harare.
- [4] Parwada C, Gadzirayi C, Karavina C, Kubiku F, Mandumbu R and Madumbu BZ. 2012. *Tagetes minuta* formulation effect *Sitophilus zeamais* (weevils) control in stored maize grain. International Journal of Plant Research, 2(3): 65-68 DOI: 10.5923/j.plant.20120203.04
- [5] Parwada C, Gadzirayi C, Karavina C, Kubiku F, Mandumbu R and Madumbu BZ. 2012. *Tagetes minuta* formulation effect *Sitophilus zeamais* (weevils) control in stored maize grain. International Journal of Plant Research, 2(3): 65-68 DOI: 10.5923/j.plant.20120203.04
- [6] Matthews GA. 1993. Insecticide application in the stores. In Matthews, GA and Hislop EC (eds.). Application technology for crop protection. CAB, London, pp. 305-315.
- [7] Giga DP, Mutemererwa S, Moya G and Neeley D. 1991. Assessment and control of losses caused by insect pests in small farmers' stores in Zimbabwe. Crop Protection, 10: 287-292.
- [8] Arex 2004. An overview of Zimbabwe's Agriculture, Ministry of Lands and Agriculture. Government Printers. Harare.
- [9] Boxall RA, Brice JR, Taylor SJ and Bancroft RD. 2002. Technology and management of storage. In: Golob P, Farrell G and Orchard JE (eds.), Crop Post-Harvest: Science and Technology, Principles and Practice. Natural Resource Institute, University of Greenwich, 1:141-232.
- [10] Matthews GA. 1993. Insecticide application in the stores. In Matthews, GA and Hislop EC

- (eds.). Application technology for crop protection. CAB, London, pp. 305-315.
- [11] Rukuni M and Eitcher CK. 1994. Agricultural Revolution. University of Zambia Publications. Pp 208-212.
- [12] Umoetok SBA, Osuagwu AN, Udo IA, Idiongette MI and Ukeh DA. 2009. Effects of Azadirachta indica products on the management of Oothea mutabilis on Telfairia occidentalis in Calabar, Southeast Nigeria. Crop Protection, 28: 583-587. <http://dx.doi.org/10.1016/j.cropro.2009.03.002>
- [13] Chaubey MK. 2008. Fumigant toxicity of essential oils from some common species against pulse beetle Callosobruchus chinensis (Coleoptera: Bruchidae). J. Oleo Sci., 57: 171-179.
- [14] Chaubey MK. 2008. Fumigant toxicity of essential oils from some common species against pulse beetle Callosobruchus chinensis (Coleoptera: Bruchidae). J. Oleo Sci., 57: 171-179
- [15] Masoka JK. 2002. Maize production facts and trends in Zimbabwe. University of Zimbabwe Publishers, Harare
- [16] Boxall RA, Brice JR, Taylor SJ and Bancroft RD. 2002. Technology and management of storage. In: Golob P, Farrell G and Orchard JE (eds.), Crop Post-Harvest: Science and Technology, Principles and Practice. Natural Resource Institute, University of Greenwich, 1:141-232.
- [17] Dent D. 1991. Insect pest management .CAB International, United Kingdom.
- [18] Derera J, Pixley KV and Giga PD. 1999. Inheritance of maize weevil resistance in maize hybrids among lines in Southern Africa, Mexico and CIMMYT – Zimbabwe. Pp. 19-23.
- [19] Keba T and Sori W. 2013. Differential resistance of maize varieties to maize weevil (Sitophilus zeamais Motschulsky) (Coleoptera: Curculionidae) under laboratory conditions. Journal of Entomology, 10(1): 1-12.
- [20] Umoetok SBA, Osuagwu AN, Udo IA, Idiongette MI and Ukeh DA. 2009. Effects of Azadirachta indica products on the management of Oothea mutabilis on Telfairia occidentalis in Calabar, Southeast Nigeria. Crop Protection, 28: 583-587. <http://dx.doi.org/10.1016/j.cropro.2009.03.002>.
- [21] Pingali PL and Pandey S. 2000. Meeting world maize need: technological opportunities and priorities for the public sector. In: 1999/2000 CIMMYT World Maize Facts and Trends, 1-9.
- [22] Tefera T, Mugo S, Tende R and Likhayo P. 2011. Methods of screening maize for resistance to stem borers and post-harvest insect pests. CIMMYT, Nairobi, Kenya.
- [23] Dent D. 1991. Insect pest management .CAB International, United Kingdom.
- [24] Dent D. 1991. Insect pest management .CAB International, United Kingdom.
- [25] Mihm, J.A. 1997. Insect Resistant Maize: Recent Advances and Utilization: Proceeding of an International Symposium Held at the International Maize and Wheat Improvement Centre (CIMMYT), 27 November – 3 December, 1994. CIMMYT.
- [26] Mihm, J.A. 1997. Insect Resistant Maize: Recent Advances and Utilization: Proceeding of an International Symposium Held at the International Maize and Wheat Improvement Centre (CIMMYT), 27 November – 3 December, 1994. CIMMYT.
- [27] Pingali PL and Pandey S. 2000. Meeting world maize need: technological opportunities and priorities for the public sector. In: 1999/2000 CIMMYT World Maize Facts and Trends, 1-9.
- [28] Tefera T, Mugo S, Tende R and Likhayo P. 2011. Methods of screening maize for resistance to stem borers and post-harvest insect pests. CIMMYT, Nairobi, Kenya.
- [30] Metcalf RL and Luckman WH. 1975. Introduction to Insect pest management. John Wiley and Sons, Canada.